



Ventilation-perfusion Scan: A Functional Imaging Approach to Regional Lung Disease in Primary Ciliary Dyskinesia

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Abstract:

Background: As a rare genetic disorder defined by defective ciliary motility, Primary Ciliary Dyskinesia (PCD) leads to chronic respiratory complications. A founder variant in the *RSPH4A* [c.921+3_921+6del] gene, common in Puerto Rico, impairs ciliary function and contributes to progressive lung disease. Today, the clinical value of ventilation/perfusion (V/Q) scans in PCD has not been thoroughly investigated, as well as pulmonary function tests (PFTs) and chest imaging.

Objective: To explore the role and clinical utility of V/Q scans in PCD.

Methods: Retrospective evaluation of pulmonary function tests, chest imaging, and Tc-99m DTPA ventilation and Tc-99m MAA perfusion scans in ten patients with genetically confirmed *RSPH4A*-associated PCD.

Results: Regional lung dysfunction was present in V/Q scans of patients with PCD. Age-related FEV₁ decline and severity of bronchiectasis on chest imaging were depicted in V/Q scans.

Conclusion: V/Q scans may identify functional abnormalities in PCD and complement standard imaging and pulmonary function tests. These scans may serve as valuable tools for monitoring disease progression and informing clinical decision-making.

Keywords: Ciliary dyskinesia, V/Q scan, *RSPH4A*, Lung Disease, Bronchiectasis, imaging.

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1. INTRODUCTION

Primary Ciliary Dyskinesia (PCD) is a rare, genetically heterogeneous disorder caused by defects in motile cilia, leading to impaired mucociliary clearance and chronic respiratory disease [1]. Neonatal respiratory distress, persistent wet cough, sinusitis, otitis media, and bronchiectasis are common findings of this disorder [2, 3]. PCD diagnosis can be a challenge, since it often requires a

clinical history with a combination of different tools, like nasal nitric oxide measurements [4], ciliary ultrastructure assessment [5], high-speed videomicroscopy [6], and genetic testing [7]. A population-based pathogenic variant in the *RSPH4A* [c.921+3_921+6delAACT] gene has been identified as a prevalent founder variant associated with PCD in Puerto Rico [8]. The intronic deletion in the *RSPH4A* gene affects the radial spoke head protein 4A,

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which is critical for proper ciliary beat function [9]. *RSPH4A*-associated PCD shows a progressive decline in pulmonary function. Over time, patients develop increasing evidence of airway damage, bronchiectasis, and hypoxemia, despite early supportive care [10]. Pulmonary function tests (PFTs) [11], impulse oscillometry [12] and chest Computed Tomography (CT) scans [13] are commonly used to monitor lung disease progression in PCD. However, there is a gap in knowledge regarding the role of nuclear imaging in PCD, particularly the ventilation/perfusion (V/Q) scan. Today, V/Q scans are used in cystic fibrosis, chronic obstructive pulmonary disease (COPD), and pulmonary embolisms to evaluate lung function and regional perfusion mismatches [14-16]. Information about the application in PCD remains limited. Understanding the anatomic and physiologic changes present on V/Q imaging may enhance PCD disease monitoring and improve clinical decisions. V/Q scans may predict the need for therapeutic escalation or transplant evaluation. Our communication tries to explore the medical relevance of V/Q scans in characterizing PCD pulmonary involvement, especially among our patients with *RSPH4A*-associated PCD in Puerto Rico.

2. MATERIALS AND METHODS

A retrospective review of clinical data from a cohort of ten ($n=10$) patients from Puerto Rico with PCD carrying the homozygous or compound heterozygous *RSPH4A* [c.921+3_921+6delAAGT] genetic variant was conducted. The ten patients were evaluated at the PCD center in Puerto Rico between 2019 and 2024. Inclusion criteria required a confirmed PCD genetic diagnosis and the availability of V/Q scans, PFTs, and chest imaging. Pulmonary function tests included spirometry (FEV₁), which were performed according to ATS/ERS guidelines [17]. V/Q scans were reviewed as part of the

comprehensive pulmonary evaluation at diagnosis to identify regional ventilation-perfusion mismatch. Tc-99m DTPA (diethylene-triamine-pentaacetate) aerosol for ventilation imaging and Tc-99m macroaggregated albumin (MAA) for perfusion were used in all V/Q scans included in this study. All V/Q scans were completed at the same clinical center and with the same equipment. A board-certified nuclear medicine physician interpreted and reviewed all scans for the presence and distribution of V/Q mismatches or defects. All chest CT scans were analyzed to assess the presence and severity of bronchiectasis, and other structural abnormalities among different ages in axial and coronal views. Correlations and comparisons between imaging findings, age, and FEV₁ were completed. The Institutional Review Board of Ponce Health Sciences University (IRB protocol #2309163682) approved this protocol on February 5, 2024.

3. RESULTS

3.1. Subjects Characteristics

A total of 10 patients ($n=10$) with genetically confirmed PCD, all homozygous for the *RSPH4A* [c.921+3_921+6delAAGT] genetic variant, were evaluated with PFTs, chest CT scans, and V/Q scans. The cohort in this study included both pediatric ($n=5$) and adult ($n=5$) patients. Age ranged from 11 to 55 years, with 7 out of 10 patients being females. Table 1 shows patient characteristics, including age, sex, genotype, and infection status.

3.2. Pulmonary Function Tests (PFTs)

Preserved lung function (mean FEV₁: 62%) was present in pediatric patients, whereas adult patients demonstrated moderate to severe airflow limitation (mean FEV₁: 43%) in most cases. A 11-year-old male with *Burkholderia cepacia* had the lowest FEV₁ (48%) among all pediatric patients.

Table 1. Clinical and demographic characteristics: Demographic data, clinical parameters, and imaging findings of ten patients with PCD. The FEV₁ shows a trend of declining values with age. *Pseudomonas aeruginosa* predominates among adults. Younger patients with PCD presented with other organisms such as *Burkholderia cepacia* and *Haemophilus parainfluenzae*. Varying degrees of bronchiectasis severity were present, with adult patients exhibiting more structural lung damage than pediatric patients. V/Q scans revealed physiologic abnormalities consistent with V/Q mismatch in all cases.

Patient ID	Age (years)	Sex	FEV ₁ (%)	Infection Status	Bronchiectasis (chest CT scan)	V/Q Scan Findings
PCD-01	11	Male	48	<i>B. cepacia</i>	Moderate	Patchy lower lobe defects
PCD-02	12	Male	66	<i>H. parainfluenzae</i>	Mild	Mild peripheral defects
PCD-03	15	Female	73	<i>P. aeruginosa</i>	Mild	Moderate mismatch
PCD-04	18	Female	66	<i>P. aeruginosa</i>	Moderate	Moderate mismatch
PCD-05	20	Female	58	<i>P. aeruginosa</i>	Moderate	Moderate mismatch
PCD-06	34	Female	39	<i>P. aeruginosa</i>	Severe	Bibasilar defects
PCD-07	40	Female	42	<i>P. aeruginosa</i>	Severe	Bibasilar defects
PCD-08	42	Female	53	<i>P. aeruginosa</i>	Severe	Bibasilar defects
PCD-09	55	Female	51	<i>P. aeruginosa</i>	Severe	Severe mismatch
PCD-10	55	Male	32	<i>P. aeruginosa</i>	Severe	Severe mismatch

3.3. Chest CT Scan

The CT scans showed the presence of worsening bronchiectasis with age, mucus plugging, and airway wall thickening, with more severe abnormalities observed in adults (Fig. 1). All pediatric patients presented early structural changes in the lower lobes. The adults demonstrated widespread bilateral bronchiectasis and architectural distortion on CT scans.

3.4. Ventilation/Perfusion (V/Q) Scan

All patients with PCD demonstrated V/Q scans with mismatches (Fig. 2). Pediatric patients showed peripheral ventilation defects, with mostly preserved perfusion. Compared with children, adults showed pronounced mismatches. The extent of V/Q abnormalities increases with age, reductions in FEV₁, and radiographic changes on chest CT scans.

4. DISCUSSION

This study explores the role of complementary pulmonary function tests, chest CT scans, and V/Q scans in a genetically homogeneous PCD cohort to create a multidimensional view of disease progression, using V/Q mismatch as a possible marker of worsening lung function. A significant V/Q mismatch as a hallmark of lung dysfunction in Puerto Rican patients with PCD carrying the homozygous *RSPH4A* [c.921+3_921+6delAAGT] genetic variant was depicted in this study. The cohort of patients with PCD included in this study exhibited functional abnormalities on V/Q scans. The adult patients showed more extensive and severe defects as compared with pediatric patients. The physiological impairments were accompanied by an age-related decline in FEV₁. Furthermore, there was increased bronchiectasis severity with age on chest CT scans.

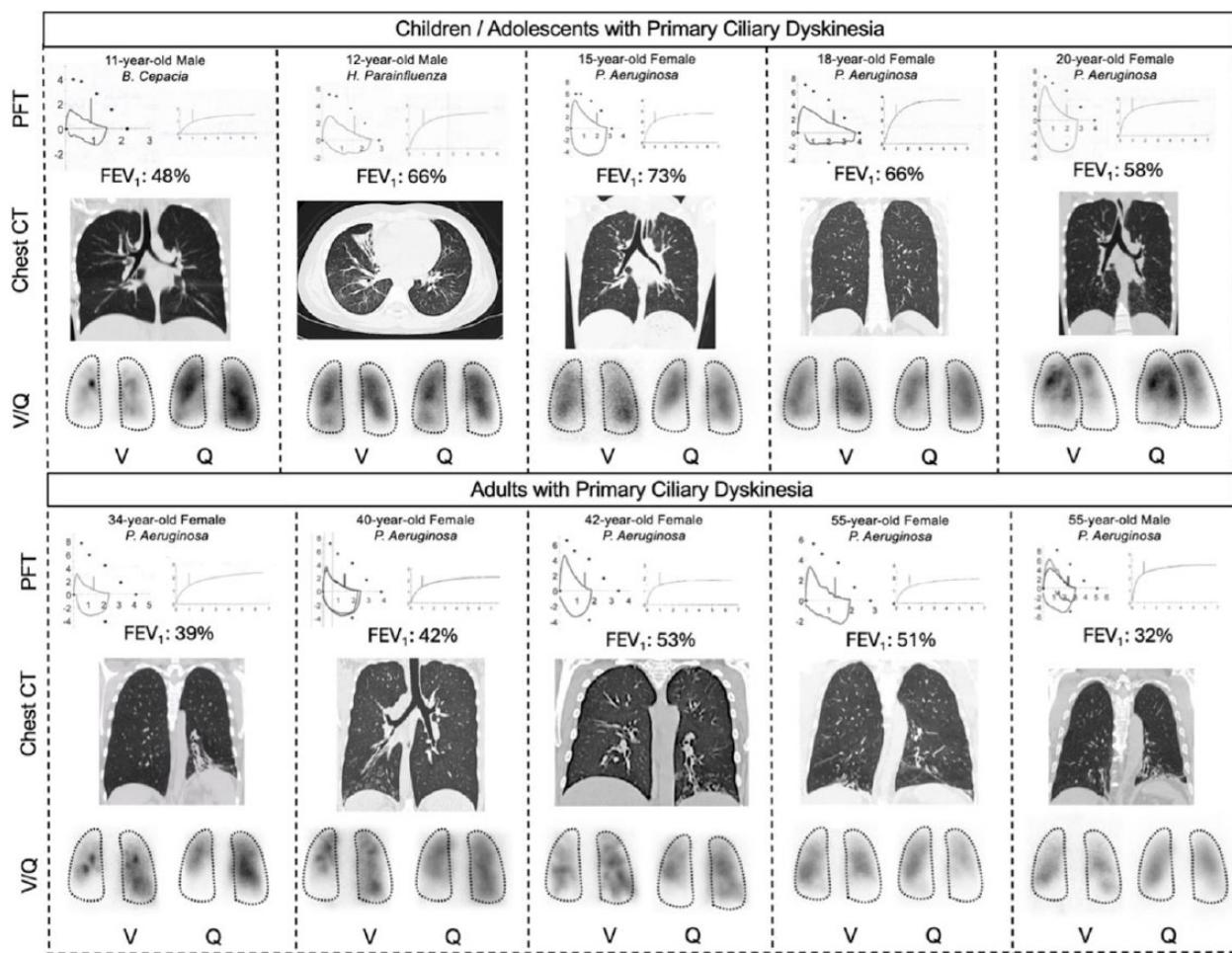


Fig. (1). Pulmonary function testing, chest CT imaging, and ventilation/perfusion (V/Q) scans in pediatric and adult patients with primary Ciliary Dyskinesia (PCD) Carrying the Homozygous *RSPH4A* [c.921+3_921+6delAAGT] genetic variant.

Each panel includes FEV₁ values, coronal or axial chest CT images, and posterior V/Q scan views. Tc-99m DTPA was used for ventilation, and Tc-99m MAA for perfusion. A progressive decline in lung function is observed across age groups, with worsening FEV₁ values and increasingly severe bronchiectasis on CT scans. V/Q scans demonstrate a corresponding increase in ventilation-perfusion defects, particularly in the lower lobes.

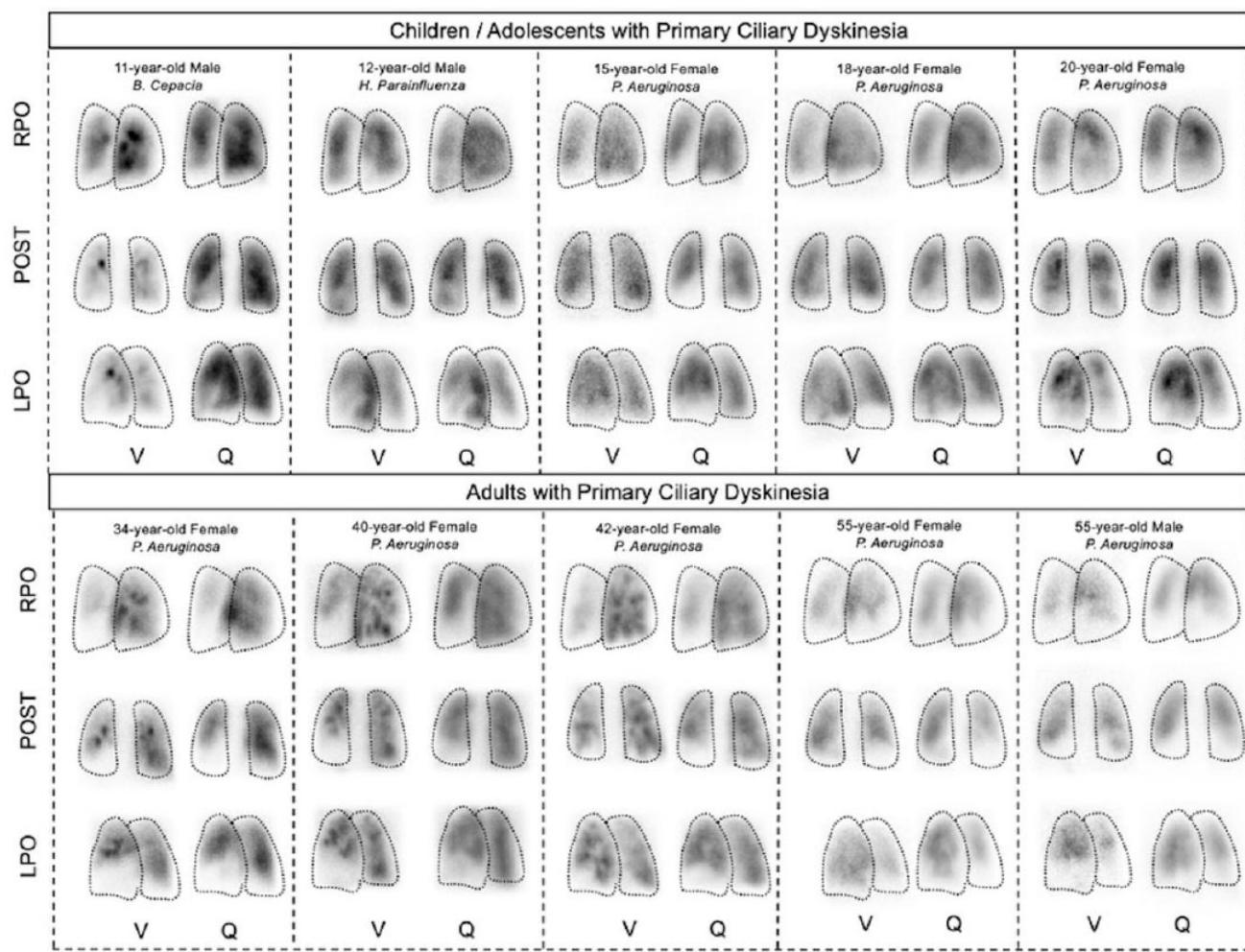


Fig. (2). Ventilation/perfusion (v/q) scans in pediatric and adult patients with primary ciliary dyskinesia (PCD) carrying the homozygous *RSPH4A* [c.921+3_921+6delAAGT] genetic variant.

Representative planar V/Q scan images from children, adolescents, and adults with genetically confirmed PCD are shown in anterior (ANT), posterior (POST), right posterior oblique (RPO), and left posterior oblique (LPO) projections.

The imaging abnormalities observed in this study reflect the underlying mechanism of PCD. The dysfunctional ciliary motility leads to chronic mucus accumulation, which increases the risk of recurrent infections, progressive airway changes, and remodeling [10]. Several mechanisms affect PCD, including mucus plugging, atelectasis, and bronchial wall damage, all of which may result in regional ventilation defects and localized hypoxic vasoconstriction, causing secondary perfusion defects [18]. These mechanisms were evident in our V/Q findings, where ventilation abnormalities appeared early in pediatric patients. The perfusion defects became more pronounced in adults with parallel structural damage seen on chest CT scans. Previous studies have shown that V/Q mismatch is a sensitive indicator of regional lung dysfunction; examples include diseases such as cystic fibrosis [19] and chronic obstructive pulmonary disease [20], in which heterogeneous lung involvement is common.

The findings in V/Q abnormalities across patients with PCD, and their correlation with age and FEV₁, support the potential role of V/Q scans as a tool for future monitoring of PCD progression. The V/Q scans may detect physiologic impairment before gross structural changes become evident on chest CT scans [21], particularly in pediatric patients. In our cohort, pediatric patients with relatively preserved spirometry already exhibited patchy ventilation defects. These findings explore the possible utility of V/Q scans in identifying early stages of disease, which aligns with MRI-based studies in PCD showing that ventilation and perfusion abnormalities can precede spirometric decline [22, 23]. The identification of regional functional impairment by V/Q scans may help PCD providers tailor personalized interventions. These may include intensifying airway clearance strategies and counseling adults with extensive V/Q defects about early referral for lung transplant evaluation.

In cystic fibrosis literature, perfusion abnormalities on V/Q scans have been shown to correlate with disease severity and prognosis, with greater disparity predicting poorer outcomes [24]. Furthermore, in COPD, V/Q scans reveal disease heterogeneity and guide surgical planification for volume reduction and other interventional therapy [25]. Nevertheless, in PCD, the use of V/Q scans is not included in recent PCD guidelines. While radioaerosol clearance studies have proven valuable for diagnosing ciliary dysfunction [26], few studies have examined the use of standard V/Q scans to evaluate lung function or progression. The findings presented in this study begin to fill this knowledge gap by characterizing typical V/Q scan patterns in genetically defined patients with PCD and demonstrating their relationship with clinical and structural markers of disease currently used in clinical practice.

This study has several notable strengths. Our study focuses on a well-defined genetic subgroup of PCD with a single pathogenic genetic variant. The uniformity presented in this study minimizes genetic heterogeneity as a confounding factor. The results allow for more precise interpretation of clinical, functional, and imaging findings of this PCD cohort. The comparison of different variables, including chest CT imaging and FEV₁, offers a complementary perspective on PCD severity. The inclusion of pediatric and adult patients across different clinical presentations provides age-related trends in pulmonary function decline. Also, structural damage and physiologic impairment, particularly in relation to chronic infection status with *P. aeruginosa* or other PCD-related pathogens was explored. This characterization enhances the relevance and interpretability of our findings within the context of genotype-specific disease progression in PCD. Apart from the sample size and the possible generalizability of the findings, the absence of semi-quantitative scoring for V/Q abnormalities and the lack of longitudinal follow-up precluded the assessment of temporal changes or treatment response at the individual level. Future studies may use quantitative methods to identify statistical differences and validate results in cohorts with PCD genetic diversity.

The possible integration of V/Q scans with advanced functional imaging modalities, such as hyperpolarized gas MRI or dynamic V/Q MRI, may offer additional physiological insights into regional lung dysfunction in patients with PCD. Each multimodal approach in physiologic studies and imaging could ultimately support the development of precision medicine strategies in PCD. The final goal should be to enable genotype-informed treatment selection and dynamic monitoring of disease progression and therapeutic response over time.

CONCLUSION

A progressive age-related physiological and anatomical lung impairment was observed in patients with PCD carrying the homozygous *RSPH4A* [c.921+3_921+6de IAGT] genetic variant. This analysis revealed a possible pattern of declining FEV₁, increasing bronchiectasis on

chest CT scans, and worsening V/Q mismatch on scans. Moreover, V/Q scans were able to identify physiological abnormalities even in children with relatively preserved spirometry. These findings highlight the potential utility of V/Q scans as a tool for regional lung dysfunction detection. V/Q scans may support the role of current imaging modalities and PFTs in the longitudinal monitoring of PCD progression. Additional prospective studies are needed to define its prognostic value and potential to guide personalized clinical management in PCD.

AUTHORS' CONTRIBUTIONS

The authors confirm contribution to the paper as follows: W.D.-R.: Conceptualization; G.R.-R.: Methodology; G.R.-R., G.R.-O.: Software; G.R.-R., G.R.-O. and W.D.-R.: Validation, G.R.-R.: Formal analysis; W.D.-R: Investigation; M.J.R.-B., W.D.-R., R.A.M.: Resources; G.R.-R.: Data curation; G.R.-R.: Writing-original draft preparation; G.R.-R., G.R.-O.: Writing-review and editing; G.R.-R., G.R.-O., W.D.-R.: Visualization; W.D.-R., R.A.M.: Supervision; W.D.-R.: Funding acquisition. All authors have read and agreed to the published version of the manuscript.

LIST OF ABBREVIATIONS

PCD	= Primary Ciliary Dyskinesia
PFTs	= Pulmonary Function Tests
V/Q	= Ventilation/Perfusion
CT	= Computed Tomography
<i>RSPH4A</i>	= Radial Spoke Head Component 4A
<i>B. cepacia</i>	= Burkholderia cepacia
<i>H. parainfluenzae</i>	= Haemophilus parainfluenzae
<i>P. aeruginosa</i>	= Pseudomonas aeruginosa

ETHICS APPROVAL AND CONSENT TO PARTICIPATE

The study was approved by the Institutional Review Board of Ponce Health Sciences University (Protocol #2309163682) on February 5, 2024.

HUMAN AND ANIMAL RIGHTS

All procedures performed in studies involving human participants were in accordance with the ethical standards of institutional and/or research committee and with the 1975 Declaration of Helsinki, as revised in 2013.

CONSENT FOR PUBLICATION

It was waived in this case due to the retrospective nature of our study

STANDARDS OF REPORTING

STROBE guidelines were followed.

AVAILABILITY OF DATA AND MATERIALS

The data that support the findings of this study are available from the corresponding author, G.R.-R., on special request.

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CONFLICT OF INTEREST

The authors declare no conflict of interest, financial or otherwise.

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REFERENCES

- [1] Wee WB, Kinghorn B, Davis SD, Ferkol TW, Shapiro AJ. Primary ciliary Dyskinesia. *Pediatrics* 2024; 153(6): e2023063064. <http://dx.doi.org/10.1542/peds.2023-063064> PMID: 38695103
- [2] Mullowney T, Manson D, Kim R, Stephens D, Shah V, Dell S. Primary ciliary dyskinesia and neonatal respiratory distress. *Pediatrics* 2014; 134(6): 1160-6. <http://dx.doi.org/10.1542/peds.2014-0808> PMID: 25422025
- [3] Goutaki M, Shoemark A. Diagnosis of primary ciliary dyskinesia. *Clin Chest Med* 2022; 43(1): 127-40. <http://dx.doi.org/10.1016/j.ccm.2021.11.008> PMID: 35236553
- [4] Shapiro AJ, Dell SD, Gaston B, et al. Nasal Nitric Oxide measurement in primary ciliary dyskinesia. A technical paper on standardized testing protocols. *Ann Am Thorac Soc* 2020; 17(2): e1-e12. <http://dx.doi.org/10.1513/AnnalsATS.201904-347OT> PMID: 31770003
- [5] Rezaei M, Soheili A, Ziai SA, et al. Transmission electron microscopy study of suspected primary ciliary dyskinesia patients. *Sci Rep* 2022; 12(1): 2375. <http://dx.doi.org/10.1038/s41598-022-06370-w> PMID: 35149751
- [6] De Jesús-Rojas W, Demetriou ZJ, Muñiz-Hernández J, et al. Advancing primary ciliary dyskinesia diagnosis through high-speed video microscopy analysis. *Cells* 2024; 13(7): 567. <http://dx.doi.org/10.3390/cells13070567> PMID: 38607006
- [7] Horani A, Ferkol TW, Dutcher SK, Brody SL. Genetics and biology of primary ciliary dyskinesia. *Paediatr Respir Rev* 2016; 18: 18-24. PMID: 26476603
- [8] De Jesús-Rojas W, Reyes De Jesús D, Nieves AM, Mosquera RA, Martinez-Cruzado JC. Primary ciliary dyskinesia: Ancestral Haplotypes analysis of the RSPH4A founder mutation in Puerto Rico. *Cureus* 2021; 13(9): e17673. <http://dx.doi.org/10.7759/cureus.17673> PMID: 34513534
- [9] De Jesús-Rojas W, Meléndez-Montañez J, Muñiz-Hernández J, et al. The RSPH4A gene in primary ciliary dyskinesia. *Int J Mol Sci* 2023; 24(3): 1936. <http://dx.doi.org/10.3390/ijms24031936> PMID: 36768259
- [10] De Jesús-Rojas W, Shapiro AJ, Shoemark A. Respiratory aspects of primary ciliary dyskinesia. *Clin Chest Med* 2024; 45(3): 717-28. <http://dx.doi.org/10.1016/j.ccm.2024.02.020> PMID: 39069333
- [11] Halbeisen FS, Goutaki M, Spycher BD, et al. Lung function in patients with primary ciliary dyskinesia: an iPCD Cohort study. *Eur Respir J* 2018; 52(2): 1801040. <http://dx.doi.org/10.1183/13993003.01040-2018> PMID: 30049738
- [12] De Jesús-Rojas W, Reyes-Peña L, Muñiz-Hernández J, et al. Bronchiectasis assessment in primary ciliary dyskinesia: A non-invasive approach using forced oscillation technique. *Diagnostics* 2023; 13(13): 2287. <http://dx.doi.org/10.3390/diagnostics13132287> PMID: 37443681
- [13] Kennedy MP, Noone PG, Leigh MW, et al. High-resolution CT of patients with primary ciliary dyskinesia. *AJR Am J Roentgenol* 2007; 188(5): 1232-8. <http://dx.doi.org/10.2214/AJR.06.0965> PMID: 17449765
- [14] Itti E, Fauroux B, Pigeot J, et al. Quantitative lung perfusion scan as a predictor of aerosol distribution heterogeneity and disease severity in children with cystic fibrosis. *Nucl Med Commun* 2004; 25(6): 563-9. <http://dx.doi.org/10.1097/01.mnm.0000126508.96340.1d> PMID: 15167515
- [15] Mortensen J, Berg RMG. Lung Scintigraphy in COPD. *Semin Nucl Med* 2019; 49(1): 16-21. <http://dx.doi.org/10.1053/j.semnuclmed.2018.10.010> PMID: 30545511
- [16] Costello P, Gupta KB. Pulmonary embolism: imaging modalities--V/Q scan, spiral (helical) CT, and MRI. *Semin Vasc Med* 2001; 1(2): 155-64. <http://dx.doi.org/10.1055/s-2001-18483> PMID: 15199498
- [17] Stanojevic S, Kaminsky DA, Miller MR, et al. ERS/ATS technical standard on interpretive strategies for routine lung function tests. *Eur Respir J* 2022; 60(1): 2101499. <http://dx.doi.org/10.1183/13993003.01499-2021> PMID: 34949706
- [18] Dunham-Snary KJ, Wu D, Sykes EA, et al. Hypoxic pulmonary vasoconstriction. *Chest* 2017; 151(1): 181-92. <http://dx.doi.org/10.1016/j.chest.2016.09.001> PMID: 27645688
- [19] Gustafsson PM, Aurora P, Lindblad A. Evaluation of ventilation maldistribution as an early indicator of lung disease in children with cystic fibrosis. *Eur Respir J* 2003; 22(6): 972-9. <http://dx.doi.org/10.1183/09031936.03.00049502> PMID: 14680088
- [20] Hwang HJ, Lee SM, Seo JB, et al. Assessment Of changes in regional xenon-ventilation, perfusion, and ventilation-perfusion mismatch using dual-energy computed tomography after pharmacological treatment in patients with chronic obstructive pulmonary disease: Visual And quantitative analysis. *Int J Chron Obstruct Pulmon Dis* 2019; 14: 2195-203. <http://dx.doi.org/10.2147/COPD.S210555> PMID: 31576116
- [21] Roach PJ, Bailey DL, Harris BE. Enhancing lung scintigraphy with single-photon emission computed tomography. *Semin Nucl Med* 2008; 38(6): 441-9. <http://dx.doi.org/10.1053/j.semnuclmed.2008.06.002> PMID: 19331838
- [22] Wucherpfennig L, Wuehnemann F, Eichinger M, et al. Magnetic resonance imaging of pulmonary and paranasal sinus abnormalities in children with primary ciliary dyskinesia compared to children with cystic fibrosis. *Ann Am Thorac Soc* 2024; 21(3): 438-48. <http://dx.doi.org/10.1513/AnnalsATS.202305-453OC> PMID: 38206973
- [23] Smith LJ, West N, Hughes D, et al. Imaging lung function abnormalities in primary ciliary dyskinesia using hyperpolarized gas ventilation MRI. *Ann Am Thorac Soc* 2018; 15(12): 1487-90. <http://dx.doi.org/10.1513/AnnalsATS.201711-890RL> PMID: 29684285
- [24] Stanchina ML, Tantisira KG, Aquino SL, Wain JC, Ginns LC. Association of lung perfusion disparity and mortality in patients with cystic fibrosis awaiting lung transplantation. *J Heart Lung Transplant* 2002; 21(2): 217-25. [http://dx.doi.org/10.1016/S1053-2498\(01\)00376-X](http://dx.doi.org/10.1016/S1053-2498(01)00376-X) PMID: 11834350
- [25] Kristiansen JF, Perch M, Iversen M, Krakauer M, Mortensen J. Lobar quantification by Ventilation/Perfusion SPECT/CT in patients with severe emphysema undergoing lung volume reduction with endobronchial valves. *Respiration* 2019; 98(3): 230-8. <http://dx.doi.org/10.1159/000500407> PMID: 31167210
- [26] Marthin JK, Mortensen J, Pressler T, Nielsen KG. Pulmonary radioaerosol mucociliary clearance in diagnosis of primary ciliary dyskinesia. *Chest* 2007; 132(3): 966-76. <http://dx.doi.org/10.1378/chest.06-2951> PMID: 17646235

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