Low Doses of Kretek Cigarette Smoke Altered Rat Lung Histometric, and Overexpression of the p53 Gene

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Abstract:
Background: The components of kretek cigarettes include tobacco as the main part, clove, and sauce. Filtered kretek cigarettes are kretek cigarettes that have one end filtered. Cigarette smoke contributes to the disruption of the respiratory system, so it is necessary to know the effect of low doses of cigarette smoke on changes in the histometric of the respiratory system, and whether it affects p53 gene expression. This study aims to determine changes in the histometric of the respiratory system and p53 gene expression.

Methods: In this study, we used Sprague-Dawley rats. Group I of rats breathing normal air, were not exposed to filtered kretek cigarette smoke (as a control). Group II of rats, as a treatment group, were exposed to filtered kretek cigarette smoke 1 stick/day for 3 months. The results of lung histometry measurements and p53 gene expression between groups were analyzed using the Independent Sample T-test. The difference between groups is significant if the test results show P < 0.05.

Results: Bronchioles length, width, area, and perimeter in group I were 40.55±1.57 μm, 14.82±0.41 μm, 494.61±5.62 μm², and 233.87±4.51 μm, respectively. Bronchioles length, width, area, and perimeter in group II were 30.76±0.78 μm, 9.28±0.40 μm, 297.32±2.53 μm², and 177.84±5.15 μm, respectively. The area and perimeter of respiratory bronchioles in group I were 17.68±0.49 μm², and 26.60±0.52 μm respectively, while those in group II were 19.28±0.35 μm², and 29.28±0.35 μm, respectively. Mucus was found in the bronchioles and respiratory bronchioles in group II, however, there was no visible mucus observed in group I. In addition, it was also concluded that exposure to low doses of filtered kretek cigarette smoke, 1 cigarette/day for 3 months, increased the expression of the p53 gene in the lungs of rats.

Conclusion: The size of bronchioles in rats decreased after being exposed to filtered kretek cigarette smoke 1 stick/day for 3 months, while the size of respiratory bronchioles increased. In addition, exposure to cigarette smoke increased the expression of the p53 gene in the rat lungs.

Keywords: Filtered kretek cigarettes smoke, Histometric, Bronchioles, Respiratory bronchioles, p53 gene.
1. INTRODUCTION

In Indonesia, many people smoke, especially among young people [1]. Cigarettes can be obtained from the market, but some are made by residents, manually. The main ingredient of cigarettes is tobacco. Cigarettes are one of the major health problems in the world. Even so, many people smoke. It has been reported that cigarettes kill more than 8 million people each year in the world. More than 7 million of those deaths are the result of direct tobacco use, while around 1.2 million are the result of non-smokers being exposed to second-hand smoke [2]. The other reports demonstrated that smoking is a risk factor for various types of diseases, because smokers should stop smoking before the age of 40 years [3]. Especially Indonesia, is one of the countries in Southeast Asia whose population is mostly smokers [4]. Moreover, the number of smokers in Indonesia is very large. In addition, filtered kretek cigarettes are in great demand by most active smokers in Indonesia. This fact shows that most Indonesians smoke kretek.

There are several types of cigarettes in Indonesia, including filtered kretek cigarettes. Generally, filtered kretek cigarettes are made in the factory using machines and are equipped with a filter at one end. In an earlier period, the kretek cigarettes were not filtered at one end. Until now, on the market circulating filtered kretek cigarettes, and non-filtered kretek cigarettes. Filtered kretek cigarettes contain tobacco (as the main ingredient) that is mixed with cloves and sauces [5]. In addition to the added cloves, there are other ingredients to complement the taste of kretek cigarettes, namely sauce.

The scientific name of cultivated tobacco is *Nicotiana tabacum* Linn. Based on a previous report, the Indonesia cigarette industry (±80%) used local tobacco such as Temanggung tobacco, Weleri tobacco, Yogyakarta tobacco, Muntial Tobacco, Boyolali tobacco, Paiton tobacco, Kasturi tobacco, and Madura tobacco [6]. The dried clove flower buds (*Syzygium aromaticum* (L.) Merr. & L.M. Perry) [7] were used as dough in filtered kretek cigarettes. Filtered kretek cigarette sauce is a unique taste of cigarettes [8]. In Klaten district and its surroundings (Central Java Province, Indonesia), kretek cigarette sauce is known as “woor”.

Cigarette smoke contains a complex mixture of chemical compounds. More than 4000 chemical compounds in cigarette smoke have been identified [9]. Cigarette smoke is known to consist of gas (92%) and solid or particulate matter (8%) [10]. Recent research demonstrates that particulate matter emissions from kretek cigarettes can be very high. This is clearly dangerous for both active smokers and non-smokers who are exposed to cigarette smoke [11]. People who smoke, gases and particulate matter in the smoke pour from the mouth to the alveoli. It has been demonstrated that there was deposition of cigarette smoke particles in various locations of the human respiratory tract [12].

Furthermore, it has been demonstrated that cigarette smoke is associated with bronchopulmonary dysplasia [13]. In addition, cigarette smoke also has free radicals that cause airway disorders, resulting in lung abnormalities. Moreover, cigarette smoke contains particles that damage lung tissue, causing permanent damage called chronic obstructive pulmonary disease (COPD) [14, 15].

Generally, a rat’s respiratory system consists of the nose, pharynx, larynx, trachea, primary bronchi, and lungs (bronchioles and alveoli) [16]. The latest research demonstrated that components of rat lung structure are blood vessels, bronchi, terminal bronchi, respiratory bronchioles, alveolar ducts, alveolar sacs, and alveoli [17]. In addition, it has been demonstrated about the lower respiratory system of mice (bronchioles, alveolar passages, and alveoli) both qualitatively and quantitatively [18]. A previous study reported that the respiratory systems of rats exposed to gas showed increased tissue elasticity and tissue resistance [19]. A recent study also reported that rats were given conventional cigarette smoke, collagen deposition appearance in the bronchioles, as well as intrabronchial mucus [20].

In addition, it is also shown that cigarette smoke affects the anatomical structures of the respiratory system [21]. Another study also showed that smokers have experienced basal cell hyperplasia, mucous cell hyperplasia, and squamous metaplasia [22]. In the application of COPD diagnosis, both qualitative and quantitative assessments are needed [23]. This research is important because bronchioles and respiratory bronchioles are an important part of the lungs associated with the alveoli. We intend to present the quantitative changes in bronchioles, and respiratory bronchioles caused by the effects of filtered kretek cigarette smoke. Quantitative changes in the bronchioles and respiratory bronchioles are important so that the diagnosis related to lung tissue disorders can be determined more objectively. Apart from that, it also needs to be proven whether low doses of kretek cigarette smoke affect gene expression, especially p53. We know that the p53 gene is used as a tumour marker as a key cellular process such as proliferation, apoptosis, and metabolism to suppress tumorigenesis [24]. Therefore, based on the description above, it is necessary to research the histometric of bronchioles, and respiratory bronchioles in Sprague-Dawley rats after treatment with filtered kretek cigarette smoke. Likewise, it is also necessary to prove the dangers of low-dose smoking on over-expression of the p53 gene.

2. MATERIALS AND METHODS

2.1. Preparation of Filtered Kretek Cigarettes

The type of tobacco used as an ingredient in filtered kretek cigarettes was local tobacco. The types of local tobacco (*Nicotiana tabacum* Linn.) werecollected from areas in Manisrenggo District, Klaten Regency, Central Java Province, and Indonesia. Cloves (*Syzygium aromaticum* Merr. & L.M. Perry) used in filtered kretek cigarette dough weredried clove flowers, collected from areas in the Logede Village, Karangnongko District, Klaten Regency, Central Java Province, and Indonesia. The sauce
known as “woor,” used for filtered kretek cigarettes was a special ingredient that was always used to mix cigarettes manually. These materials were used to make filtered kretek cigarettes manually.

2.2. Ethical Clearance

The ethical clearance of this research was obtained from the Research Ethics Commission, Faculty of Medicine, Universitas Trisakti, Jakarta, Indonesia (No. 184/KER-FK/VIII/2018).

2.3. Flowchart of Experimental Rat Treatment

The flowchart of experimental rat treatment is presented in Fig. (1).

2.4. Animal Care

We used *Rattus norvegicus*, Sprague-Dawley strain. The number of rats per group was calculated using formula \((n-1) (t-1) \geq 15\). For each treatment group a minimum of 8 rats were required. Sixty male rats were used in this study. The rats included in this study were 2-3 months old and weighed 150-200 grams. Determination of rat health was carried out after examination by a Veterinary. Rats were caged (individually) in an air-conditioned room and the room temperature was set to \(22\pm3\) °C, humidity 55±5%, and artificial fluorescent lamps (12:12 hour light and dark cycle). The rats fed and drank in the libitum according to the standard. There are 2 stages in this research, namely the 1st stage in the form of rat acclimatization and the 2nd stage was the treatment of giving filtered kretek cigarette smoke to rats.


2.5. Treatment on Sprague-dawley Rats

Trial of filtered kretek cigarette smoke on rats was carried out at The Veterinary Medical Teaching Hospital (Rumah Sakit Hewan Pendidikan=RSHP), Faculty of Veterinary Medicine of Bogor Agricultural University. This research was conducted between November 2019 - April 2020.
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Fig. (2). Schematic diagram of filtered kretek cigarettes smoke exposure to a groups of Sprague-Dawley rats. A. Filtered kretek cigarettes burned. B. Tube to accommodate filtered kretek cigarettes smoke. C. Electric pump. D. Glass box for rats treatment with length 40 cm, width 30 cm, and height 20 cm).

Acclimatization of rats was carried out for 2 weeks, then continued with treatment according to groups. Rats were divided into 2 groups with randomization. Group Iqwas the rat group that was breathing normal air, and was not exposed to filtered kretek cigarette smoke (as a control). Group II, namely the rat group exposed to filtered kretek cigarettes, smoke 1 stick/day/group (Fig. 2). The treatment duration for the two groups of rats was 3 months. After the treatment, the rats were sacrificed, and then their respiratory organs were collected from the trachea to the lungs to make histology slides.

2.6. Measurement of Bronchioles and Respiratory Bronchioles

The tissue was immersed in a neutral buffer solution of 10% formalin for 24 hours at room temperature. The tissue was cut to a size of 1×1×1 cm, and then put in a tissue cassette. The tissue was then transferred to alcohol dehydration with alcohol concentrations of 70%, 80%, 90%, and 96%, respectively. The dehydration time was 2 hours for each alcohol concentration. The next stage was clearing, after which, the tissue was ready to be inserted into the paraffin block. Furthermore, embedding and blocking were carried out. The tissue in the paraffin block was cut using a microtome with a thickness of 4-5 μ, and then stained with hematoxylin and eosin.

Image documentation using Optilab Advance Plus and Image Raster 3 by PT MICONOS, Daerah Istimewa Yogyakarta, Indonesia; available at https://miconos.ac.id/new/support/download. Optilab Advance Plus and Image Raster 3 programs were used to analyze rat respiratory system histometrics (bronchioles and respiratory bronchioles) by three observers. Data for all groups were expressed as mean± standard deviation. Analyzing data between groups using an independent sample t-test. The difference between groups was significant if the test results showed P<0.05.

2.7. Expression of p53 Gene in Rat Lung

Measurement of p53 gene expression was carried out using real-time polymerase chain reaction (RT PCR). Lung tissue samples of Sprague Dawley rats were isolated for mRNA, and then the solution was put into a 1.5 mL tube, stored at –80 °C. The reagents for measuring mRNA levels were Bioline SensiFast Sybr Lo-ROX One-step kit, Zymo Research Quick-RNA™ MiniPrep Plus, and Geneaid Micropestle. The p53 rat target gene used the sense primer (5’ to 3’) AGGCCTTGGAACTCAAGGAT, while the antisense primer (5’ to 3’) TGAGTCAGGCCCTTCTGTCT, with a size of 140 bp. The glyceraldehyde-3-phosphate

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<th>Table 1. Comparison of the effect of filtered kretek cigarette smoke on the Sprague Dawley rat lung.</th>
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Abbreviations: Group I, rats breathe using ordinary air without exposure to filtered kretek cigarette smoke. Group II, the group of rats exposed to filtered kretek cigarette smoke 1 stick/day for 3 months of treatment.
dehydrogenase (GAPDH) gene was used as an internal control. The sense primer of GAPDH gene was (5' to 3') CCAGGTGGTCTCCTCTGACTTCTC, while the antisense primer was (5' to 3') ATACCAGGAAATGAGCTTGACA, with a size of 147 bp [25]. RT PCR analysis using SYBR Green Master Mix (Bio-rad) in the Bio-rad CFX96 RT PCR system. Ct p53 gene was normalized with Ct gene GAPDH. The calculation was based on the $2^{-\Delta\Delta C_t}$ method (Livak method) [26], as demonstrated in Table 1.

3. RESULTS

3.1. Filtered Kretek Cigarettes

The constituent materials of filtered kretek cigarettes are presented in Fig. (3). Filtered kretek cigarettes consist of filters and kretek dough. Tobacco is the main ingredient of kretek cigarettes, which is mixed with cloves and sauce.


Fig. (4). Hair of the Sprague-Dawley rats. A. Hair appearance of the back body. B. Hair appearance of the abdomen. I. Group I, rats breathe using ordinary air without exposure to filtered kretek cigarette smoke. II. Group II, the group of rats exposed to filtered kretek cigarette smoke 1 stick/day for 3 months of treatment.
3.2. Hair Appearance

The hair appearance of rats between group I and group II is presented in Fig. (4). The hair on the back and abdomen of rats in group I looks white, clean, and dense, while in group II, it looks dull yellow, less clean, and sparse.

3.3. Bronchioles Histometric

Histological of bronchioles in Sprague-Dawley rats between group I and group II is presented in Fig. (5). Treatment of filtered kretek cigarettes smoke 1 stick/day for 3 months causes mucus in the bronchioles space of rats in group II, while in group I, it is not found. In addition, the size of the bronchioles space also appears to be shrinking.

A comparison of bronchioles histometric in Sprague-Dawley rats between group I and group II is presented in Fig. (6). Based on Fig. (6), seen that bronchioles length, width, area, and perimeter in group I are different compared to group II (P=0.000). Bronchioles histometric of rats in group II <1 (P=0.000). In this study, the group of rats exposed to filtered kretek cigarette smoke had changes in bronchioles histometric, namely the decrease in length, width, area, and perimeter. The bronchioles length of rats in group II decreased by 24.14% compared to group I, while the bronchioles width of rats in group II decreased by 37.38% compared to group I. The bronchioles area of rats in group II decreased by 39.89% compared to group I, while the bronchioles perimeter of rats in group II decreased by 23.96% compared to group I. Rats in group II exert a negative response against filtered kretek cigarette smoke in the form of a decrease in the size of bronchioles histometric.
Fig. (6). Comparison of bronchioles histometric in Sprague-Dawley rats between group I compared to group II, staining with hematoxylin and eosin, objective 10x. A. Length and width of the bronchioles. B. Area and perimeter of the bronchioles. I. Group I, rats breathe using ordinary air without exposure to filtered kretek cigarette smoke. II. Group II, the group of rats exposed to filtered kretek cigarette smoke 1 stick/day for 3 months of treatment. The red arrows showed mucus.

3.4. Respiratory Bronchioles Histometric

The histological of respiratory bronchioles in Sprague-Dawley rats between group I and group II is presented in Fig. (7). Based on Fig. (7) treatment of filtered kretek cigarettes smoke 1 stick/day for 3 months causes mucus in the respiratory bronchioles space of Sprague-Dawley rats in group II, while in group I is not found.

Fig. (7). The histological of respiratory bronchioles in Sprague-Dawley rats between group I compared to group II, stained with haematoxylin and eosin, objective 10x. The yellow arrows showed mucus. RB= respiratory bronchioles, AD= alveolar ducts, A=alveoli. I. Group I, rats breathe using ordinary air without exposure to filtered kretek cigarette smoke. Group II, the group of rats exposed to filtered kretek cigarette smoke 1 stick/day for 3 months of treatment.
The area and the perimeter of respiratory bronchioles in the Sprague-Dawley rats between group I and group II is presented in Fig. (8). Fig. (8) demonstrates that the area and the perimeter of respiratory bronchioles of rats in group I < group II (P=0.000). In this study, the group of rats exposed to filtered kretek cigarette smoke had changes in respiratory bronchioles histometric, namely the increase in area and perimeter. The area of respiratory bronchioles of rats in group II increased by 9.05% compared to group I, while the perimeter of respiratory bronchioles of rats in group II increased by 10.075% compared to group I. Rats in group II had a positive response against filtered kretek cigarette smoke in the form of an increase in the area and perimeter of respiratory bronchioles.

3.5. Expression of p53 Gene

Optimization of RT PCR for the determination of mRNA levels for p53 is presented in Fig. (9). Based on the measurement data, it was demonstrated that the treatment of rats exposed to filtered kretek cigarette smoke 1 cigarette/day for 3 months influenced p53 gene expression. There was an increase in p53 gene expression in rats exposed to filtered kretek cigarette smoke. It was proven that the mRNA levels of group II were higher (upregulation) than group I (P=0.000).

4. DISCUSSION

The color of the filter on filtered kretek cigarettes before burning was different from after burning. Before the filtered kretek cigarettes were burned, the filter was white, while after they were burned, the filter had a burlywood color. It has been demonstrated that the function of the filter at the end of the filtered kretek cigarettes is to reduce tar and nicotine levels that enter the respiratory tract. The color change of the burlywood filter in this study indicated the presence of tar and nicotine from filtered kretek cigarettes. This fact is in line with previous research [27].

Filtered kretek cigarettes that were burning produced cigarette smoke and ash. Cigarette smoke was inhaled by rats, while ash of filtered kretek cigarettes was disposed of. Most of the filtered kretek cigarette smoke entered the respiratory system of rats. Filtered kretek cigarette smoke is one of the incomplete combustion products of tobacco, cloves, and sauce. Cigarette smoke includes main cigarette smoke and side cigarette smoke. Main cigarette smoke is cigarette smoke that was inhaled by rats as a model of active smokers. Side cigarette smoke is cigarette smoke that was in the environment around rats. In this study, filtered kretek cigarettes burned, and a pump mainly inhaled cigarette smoke and then channeled into the smoking box. This is similar to the smoking process that is generally carried out by smokers. The results of this study clearly show that there are differences in the color of the filters before and after the filtered kretek cigarettes burned. This fact clarifies the filter function of the filtered kretek cigarettes, which is to filter particles in cigarette smoke that are inhaled by smokers. Therefore, the filter on filtered kretek cigarettes can reduce the
Fig. (9). Optimization of RT PCR for p53 gene expression. **A.** Melting curve (derivative curve); **B.** Melting curve (normalized reporter).
content of substances inhaled when smoking. Substances in cigarette smoke that have been shown to be high in content include nicotine, various metals [28], free radicals [29], oxidants and antioxidants [30]. Even though there is a filter, cigarette smoke in this study was proven to affect the hair appearance of rats, which worsened hair appearance.

Exposure to filtered kretek cigarette smoke in this study caused the hair of rats to change from pure white to dull yellow. The hair discoloration occurred throughout the rat's body. The results in this study were in accordance with the results of the report which stated that the smoker's mustache hair turned yellow [31]. Filtered kretek cigarette smoke in this study mostly comes from tobacco leaves, so it has nicotine. Nicotine in the filtered kretek cigarette smoke enters the body and then causes discoloration, cleanliness, and density of hair in the rats. This fact is in accordance with the results of earlier studies, which show that the nicotine content of hair is used as a biomarker to assess chronic exposure to environmental tobacco smoke [32]. Nicotine in cigarette smoke by inhalation enters the body and penetrates cell membranes to the systemic circulation system. It is further stated that nicotine is the main constituent of tobacco. Nicotine is not carcinogenic, but it triggers many carcinogens in tobacco [33]. Previous research has demonstrated that nicotine levels in the hair of smokers are much higher compared to non-smokers [34]. In addition to nicotine content, cigarette smoke also causes oxidative stress that affects the condition of the hair and, among others, affects the hair fiber ([35]). In addition to affecting hair color, cigarette smoke also affects histometrics in the lungs of rats, including bronchioles and respiratory bronchioles.

The reduction in length, width, area, and perimeter of bronchioles in rats in this study responded to unwanted cigarette smoke exposure because, in normal conditions, the respiratory air does not contain cigarette smoke. This occurs as a defense mechanism for reducing the volume of cigarette smoke exposure. Reduction of bronchioles histometric reduces exposure to filtered kretek cigarette smoke, next to reducing the volume of cigarette smoke until it reaches the alveolar sacs (alveolar sacules). This situation allows obstruction of airflow that cannot flow back completely. This fact is consistent with earlier studies showing that the bronchioles are the site of airway obstruction. In addition to bronchioles abnormalities due to cigarette smoke, it was found that cigarette smoke also causes COPD [36]. Based on the facts obtained, we agree that pulmonary histometrics is useful for quantitatively measuring the tissues that make up the organs in the respiratory system. Moreover, it has been demonstrated that exposure to filtered kretek cigarettes at a dose of 1 cigarette/day or 2 cigarettes/day for 30 days, causes bronchial mucosal hyperplasia and bronchoconstriction in male Sprague Dawley rats [37]. Apart from that, it has also been demonstrated that filtered kretek cigarette smoke not only impacts the respiratory organs but also impacts Purkinje cells and pyramidal cells in Sprague Dawley rats [38].

In addition, the facts in this study, in accordance with the results of other studies which, indicated that structural abnormalities and blockage by mucus can cause destabilization of the airways in the bronchioles [39]. Previous studies have shown that morphometric measurements of rat lungs have been carried out [17]. In addition to the morphometric analysis of the rat lungs, pulmonary histometrics have also been shown. The results of the study indicated that the diameter of the respiratory bronchioles, alveolar ducts, and alveoli were 24.93±1.27 μm, 21.14±0.66 μm, and 12.95±0.21 μm, respectively [18]. Need to note that it is not only cigarette smoke that causes chronic obstruction of the bronchioles. It has been shown that chronic obstruction of the bronchioles can also be caused by cell degeneration in the bronchioles [40]. Therefore, it is still necessary to prove the effect of cigarette smoke on cell death in the bronchioles.

The effect of filtered kretek cigarette smoke in this study caused the area and perimeter of respiratory bronchioles in group II of rats to be greater compared to group I as a control. The effect of filtered kretek smoke in this study narrowed the bronchioles space (Fig. 6) but widened the respiratory bronchioles space (Fig. 8). The increased respiratory bronchioles space may be due to the pressure of filtered kretek cigarette smoke, such as the alveolar sac. The changes in histometric in the respiratory bronchioles in this study are in line with the other studies showing significant changes in the respiratory bronchioles of rats after treatment with hypoxia and melatonin [41].

Our results showed that mucus appeared in the bronchioles and respiratory bronchioles of rats. Filtered kretek cigarette smoke 1 cigarette/day for 3 months caused mucus in the respiratory bronchioles space in group II, while in group I (control), this mucus was not found (Figs. 5, 6, 7, 8). Of note, apart from in the bronchioles and respiratory bronchioles, mucus is also found in the saccus alveolar space (data not shown). The mucus is a complex mixture of mucins. Mucins are produced by airway epithelial cells and submucosal glands [42]. As a result of mucus that has accumulated in the bronchioles and respiratory bronchioles, airflow in the lungs is limited, thus resulting in lung disorders. In addition, demonstrated that inflammatory cells produce excess mucus which reduces lung function [43].

Previous studies demonstrated that cigarette smoke causes oxidative stress due to an imbalance between oxidants and antioxidants. Furthermore, cigarette smoke also induces epithelial cells for mucus hypersecretion, increasing proinflammatory cytokines and chemokines. Cigarette smoke has also been shown to increase macrophages and neutrophils and disrupt the balance of lymphocytes [30]. Cigarette smoke causes mucus hypersecretion and COPD. In addition, exposure to cigarette smoke can worsen COPD [44]. The results of other studies have also demonstrated that smoking at a young age is a dominant factor in the cause of COPD [45]. Based on the results of this study and several other studies, it is clear that cigarette smoke has a complex effect on lung tissue [46]. Moreover, cigarette smoke induces epigenetic changes in lung tissue [47], lung cancer [48, 49], oral disease, myocardial infarction, oral cancer, chronic health disease, cardiovascular disease, and asthma [50].
The Ct of the p53 gene for group I is 1, while the Ct p53 gene for group II shows a multiple of 1, so it experiences upregulation [51]. Of the 8 samples in this study, all showed up-regulation, and none showed downregulation. Rats exposed to filtered kretek cigarette smoke showed increased lung mRNA levels of the p53 gene. The occurrence of up regulation of the p53 gene in mice exposed to low doses of filtered kretek cigarette smoke in this study is in accordance with the statement relating to the processes of proliferation, apoptosis, and metabolism to suppress tumorigenesis [24]. It was further stated that the p53 gene plays an important role in responding to various types of stress cells [52]. The results of this study can be used as a basis for further research on tobacco consumption (cigarettes) with the occurrence of lung cancer. This is in accordance with research results, which demonstrate that tobacco use and variations in the p53 gene increase the risk of oral cancer [53, 54].

5. LIMITATIONS

The present study has limitations; among others, we did not measure the levels of chemical compounds in filtered kretek cigarette smoke. In this study, we have not determined damage to the bronchioles and respiratory bronchioles based on histometrics due to exposure to filtered kretek cigarette smoke. In addition, we have not performed cytométrics on bronchioles and respiratory bronchioles as we did before with human cervical cells [54, 55]. We hope that in the next opportunity, we can demonstrate the relationship between the levels of chemical compounds in filtered Kretek cigarette smoke with the level of the bronchioles and respiratory bronchioles damage based on cytometric and histometric.

CONCLUSION

Based on the results obtained, we concluded that Sprague-Dawley rats in group II were exposed to 1 stick/day of filtered kretek cigarette smoke for 3 months and showed dull yellow hair on the back and abdomen, less clean, and less frequent compared to the group I (control). The size of bronchioles of Sprague-Dawley rats decreased after being exposed to filtered kretek cigarettes smoke 1 stick/day for 3 months, while the size of respiratory bronchioles increased. Mucus is found in the bronchioles and respiratory bronchioles of Sprague-Dawley rats. Mucus was found in the bronchioles, and respiratory bronchioles in the group were exposed to 1 stick/day of filtered kretek cigarette smoke for 3 months, while in the control group, there was no visible mucus. It is clear that the quantitative changes in bronchioles and respiratory bronchioles are caused by the effects of filtered kretek cigarettes smoke. Quantitative changes in the bronchioles and respiratory bronchioles are important so that the diagnosis related to lung tissue disorders can be determined more objectively. In addition, it was also shown that exposure to low doses of filtered kretek cigarette smoke, 1 cigarette/day for 3 months, increased the expression of the p53 gene (upregulation) in the rat lungs.

SIGNIFICANCE STATEMENT

This study discovered that exposure to low-dose filtered kretek cigarette smoke changed the histometric of bronchioles and respiratory bronchioles in the Sprague-Dawley rats model. In addition, exposure to low-dose filtered kretek cigarette smoke also showed an increase in p53 gene expression in rat lungs.

ETHICS APPROVAL AND CONSENT TO PARTICIPATE

The ethical clearance of this research was obtained from the Research Ethics Commission, Faculty of Medicine, Universitas Trisakti, Jakarta, Indonesia (No. 184/KER-FK/VIII/2018).

HUMAN AND ANIMAL RIGHTS

This study adheres to internationally accepted standards for animal research, following the 3Rs principle. The ARRIVE guidelines were employed for reporting experiments involving live animals, promoting ethical research practices.

CONSENT FOR PUBLICATION

Not applicable.

AVAILABILITY OF DATA AND MATERIAL

All the data and supporting information is provided within the article.

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CONFLICT OF INTEREST

The authors declare that they have no competing interests.

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